Spring 2020 D.D. Ackerly

March 18, 2020

Readings

Hansen, T. 1997. Stabilizing selection and the comparative analysis of adaptation. Evolution 51:1341-1351.

- Ackerly, D. D. 2009. Conservatism and diversification of plant functional traits: Evolutionary rates versus phylogenetic signal. Proceedings of the National Academy of Sciences USA 106:19699-19706.
- Blomberg, S. P., and T. Garland, Jr. 2002. Tempo and mode in evolution: phylogenetic inertia, adaptation and comparative methods. J. Evol. Biol. 15:899-910.

Other strongly recommended readings!

- Blomberg, S. P., T. Garland, and A. R. Ives. 2003. Testing for phylogenetic signal in comparative data: Behavioral traits are more labile. Evolution 57:717-745. (Focus on pp. 719-723 on methods for measuring phylogenetic signal)
- Harmon, L. J., J. B. Losos, T. J. Davies, et al.. 2010. EARLY BURSTS OF BODY SIZE AND SHAPE EVOLUTION ARE RARE IN COMPARATIVE DATA Evolution 64:2385-2396.

I. Phylogenetic signal

Pattern and Process

Pattern is not process. No matter how many times we hear and repeat this phrase, the temptation to infer the action of specific processes from pattern alone is inevitable. In comparative biology, it has been all to common to equate stasis (pattern) with 'inertia' or 'constraint' (process?) and evolutionary change (pattern) with the action of selection (process). It is essential at the outset to see these dichotomies of pattern and process as orthogonal, setting up a 2x2 table:

Evolutionary process	Stasis	Change
Adaptive	Stabilizing selection Fluctuating directional selection	Directional or disruptive selection
Non-adaptive	Lack of genetic variation (= constraint?) Antagonistic correlations among traits under selection Swamping by gene flow	Mutation Genetic drift Genetic correlations with traits under selection

Pattern of trait evolution

In this case, I equate the term 'adaptive' with patterns resulting from the action of selection. It is not a statement about the functional significance of a particular trait, nor is it the same as stating that a particular trait represents an 'adaptation'. More on that in the next lecture.

It is critical to remember, as well, that these processes are neither exclusive nor independent of each other. For example, the action of drift and selection in small populations may simultaneously contribute to changes in gene frequency, and it may be difficult or impossible in an individual instance to separate their contributions. Replication in space (populations) or time (across generations) is critical, as selection is the only process we currently know of in evolutionary biology that can cause repeated, heritable change in the same direction in response to environmental conditions. Note also that selection leads to the loss of genetic variation. Thus, strong selection on a trait may lead to fixation of genes responsible for variation, such that future maintenance of the trait is due to the absence of sufficient variation (even if the selective context shifts).

As discussed in the Blomberg and Garland paper, there is a long history of usage of terms such as phylogenetic inertia or constraint, and other more recent terms such as phylogenetic effect and conservatism. To a greater or lesser degree, all of these terms imply the action of some underlying process. The term *phylogenetic signal* is useful as the implication is clear that we are only speaking about the pattern. I also like phylogenetic conservatism, though it is now taking on its own intellectual baggage and may suggest particular hypotheses about process (e.g., see Losos 2008 Ecology Letters).

Quantifying phylogenetic signal for continuous traits

Several statistical have been developed to quantify phylogenetic signal, and test for significance relative to a null model of no signal. Freckleton et al. (200x) discuss the λ parameter, which is derived from Pagel's model fitting methods. I have worked more with Blomberg et al. (2003) K statistic. λ and K have the same general interpretation:

λ, K < 1	less signal than expected under Brownian motion
λ, K = 1	expectation under B.M.
K > 1	more signal than expected under B.M.

Quantitatively, one of the main differences is that λ has a maximum around 1.1 or 1.2, so it has little resolution to describe patterns with a high degree of signal (Freckleton et al. don't report values > 1). K on the other hand has a theoretical maximum of infinity and can provide some insight into differing degrees of signal greater than expected under Brownian motion.

Fortunately, there is a simple graphical interpretation of these statistics, based on a plot of the difference in trait values between species pairs (all pairwise comparisons, not just sister taxa!) versus the phylogenetic distance between the taxa (Figure on next page). Remember from Brownian motion, the expectation of the squared difference between trait values (the variance) goes up linearly with the phylogenetic distance. It turns out that this translates approximately into a filled half triangle in a plot of the absolute difference vs. phylogenetic distance for all species pairs (assuming the tree is ultrametric). Concave curvature of the upper surface of this plot means that close relatives are less similar than expected, compared to distant relatives (K < 1). Convex curvature, or an absence of points in the lower right, means that close relatives are more similar than expected (K>1). The details of the plot structure will depend on tree topology, but in general examining these plots is quite informative. Significance tests for the correlations in such plots must be tested using Mantel tests for matrix correlations (another topic!).



Another approach to asking about phylogenetic signal and patterns is to ask whether the trait data fit an alternative model of trait evolution, and whether the parameters of this model then shed light on the processes influencing the trait and its evolutionary history. One of the very important alternative models is to introduce stabilizing selection, i.e. a model in which there is a 'pull' towards some central optimal value (which may fluctuate along different lineages), and Brownian motion reflects the random processes and/or the excursions in the action of selection around this underlying optimum. The stabilizing selection model is known as an Ornstein-Uhlenbeck stochastic process, so you will see references to OU models (and there is an R library called 'ouch' for OU models).

Another graphical approach is to draw a 'traitgram', a phylogeny where the tips are arranged left to right based on species trait values, and internal nodes show the maximum likelihood reconstructions. This can get a bit messy, as the branches cross over each other. Lots of crossovers shows that a trait has exhibited many reversals of trait evolution during its history. If those crossovers occur particularly among the most recent divergence events, then it's evidence of rapid evolution and a departure from Brownian motion, as the trait is not continuing to diverge at constant rates. Figure below is from Ackerly (2009).

As you can see from this example, the Brownian Motion model predicts a certain pattern of relative similarity – how similar are close relatives compared to the similarities among distant relatives. It is not a statement about the absolute rate of evolution, and how much the actual trait values have diverged over time. You can rescale the trait axes in the figure below so each clade spans much less or more much actual trait space, and than doesn't change the fit of the data to Brownian Motion. This is because of the unbounded assumption of the model – evolving more quickly does not result in more convergence and loss of phylogenetic signal – and in this regard continuous and discrete characters are very different.

As we discussed earlier, the variance of the Brownian Motion process is a direct measure of evolutionary rate – how quickly are trait values diverging from each other over time. Ackerly (2009) proposed using this rate parameter with standardized measures of trait variation – using log-transformed data – and branch lengths scaled in millions of years as a measure of the rate of character evolution based on comparative data (without fossils).

There are many other possible models that one can simulate and use to fit empirical trait distributions for continuous characters, which have received more or less attention in the literature.

Mode	Model	Verbal explanation
Brownian	$x_{t+1} = x_t + N(0,s)$	random walk
Brownian + trend	$x_{t+1} = x_t + N(t,s), t \neq 0$	random walk with a trend
Bounded Brownian	$x_{t+1} = x_t + N(0,s)$	random wak with lower and/or
	if $x_{t+1} < \min X$ or $> \max X$, resample	upper limits to trait values
Proportional	$x_{t+1} = x_t * LN(\log mean = 0, \log sd = s)$	multiplicative random walk
		(equivalent to a random walk on
		log of trait)
ACDC	$x_{t+1} = x_t + N(0,s)$	random walk with an accelerating
	$s = s_0 \gamma^{-t}$	or decelerating rate parameter
	$\gamma < 1$, accelerating	
	$\gamma > 1$, decelerating	
OU-1 (Ornstein-	$x_{t+1} = \mu + \theta(x_t - \mu) + N(0,s)$	stabilizing selection; random walk
Uhlenbeck)	$\mu = \text{trait optimum}$	with a 'pull' towards the optimum
	$\theta = 1$: brownian	for the clade; theta is strength of
	$\theta = 0$: complete stabilizing selection	the pull
OU-2+ (Ornstein-	$x_{t+1} = \mu + \theta(x_t - \mu) + N(0,s)$	stabilizing selection; random walk
Uhlenbeck)	$\mu = \text{trait optimum on branch i}$	with a 'pull' towards different
	$\theta = 1$: brownian	optima on different branches, to
	$\theta = 0$: complete stabilizing selection	test a priori hypotheses of adaptive
		evolution
Speciational	Brownian, with change only at time	random walk, but only one change
	of speciation	per speciation event (i.e. branch
		length = 1 between each speciation
		event)
Punctuational	Brownian, with change only at time	like speciational, but change only
	of speciation and only in one	occurs in one daughter lineage and
	daughter lineage	the other exhibits stasis

There are many evolutionists who believe that stabilizing selection is the overarching cause of evolutionary stasis and the maintenance of similarity among close relatives. The OU model has been termed a stabilizing selection model, because trait values are 'pulled' back towards an optimum value. Paradoxically, an OU process with a single optimum generates traits with K<1, so stabilizing selection reduces phylogenetic signal, which is not intuitive at first! Estes and Arnold (2007, American Naturalist) offer an important discussion of stabilizing selection and apply it to a large data set compiled by Gingerich (1983, Science) on rates of morphological evolution (mostly from fossils, not comparative data).

OU models can be used in a very different way when we have an a priori hypothesis that evolution has favored different trait values in different clades, or on different branches of a tree.

These are two-rate (or more) OU models that can use a nested-likelihood ratio approach to test whether a model with two (or more) different optima are significantly better than one optimum. This is a powerful alternative to independent contrasts, because it doesn't focus on divergence between sister taxa; rather, the focus is on the degree of similarity among taxa hypothesized to be experiencing similar adaptive regimes. We will look at the first example (Hansen 1997).

Literature:

- Abouheif, E. 1999. A method for testing the assumption of phylogenetic independence in comparative data. Evol. Ecol. Res. 1:895-909.
- Ackerly, D. D. 1999. Phylogeny and the comparative method in plant functional ecology. Pages 391-413 in M. Press, J. Scholes, and M. G. Barker, editors. Physiological plant ecology. Blackwell Scientific, Oxford, United Kingdom.
- Ackerly, D. D., and M. J. Donoghue. 1995. Phylogeny and ecology reconsidered. J. Ecol. 83:730-732.
- Ackerly, D. D., and M. J. Donoghue. 1998. Leaf size, sapling allometry, and Corner's rules: a phylogenetic study of correlated evolution in maples (<u>Acer</u>). Amer. Nat. **152**:767-791.
- Ackerly, D. D. 2009. Conservatism and diversification of plant functional traits: Evolutionary rates versus phylogenetic signal. Proceedings of the National Academy of Sciences USA 106:19699-19706. doi: 10.1073/pnas.0901635106.
- Ackerly, D.D. 2009. Phylogenetics and comparative methods. Pages 117-125 in S. Levin, ed. Guide to Ecology. Princeton Univ. Press.
- Ackerly, D.D. 2009. Phylogenetic methods in ecology. Encyclopedia of Life Sciences (ELS), John Wiley & Sons, Chichester, <u>http://www.els.net</u>. [DOI: 10.1002/9780470015902.a0021223]
- Archie, J. 1989. Homoplasy excess ratios: New indices for measuring levels of homoplasy in phylogenetic systematics and a critique of the consistency index. SystZool. **38**:253-269.
- Archie, J. 1996. Measures of homoplasy. Pages 153-188 *in* M. J. Sanderson and L. Hufford, editors. Homoplasy: the recurrence of similarity in evolution. Academic Press, San Diego.
- Blomberg, S. P., and T. Garland, Jr. 2002. Tempo and mode in evolution: phylogenetic inertia, adaptation and comparative methods. J. Evol. Biol. 15:899-910.
- Blomberg, S. P., T. Garland, Jr, and A. R. Ives. 2003. Testing for phylogenetic signal in comparative data: behavioral traits are more labile. Evolution **57**:717-745.
- Burt, B. B. 2001. Evolutionary stasis, constraint and other terminology describing evolutionary patterns. Biol. J. Linn. Soc. **72**:509-517.
- Coddington, J. 1994. The roles of homology and convergence in studies of adaptation. Pages 53-78 *in* P. Eggleton and R. Vane-Wright, editors. Phylogenetics and ecology. Academic Press, London.
- de Queiroz, K. 1996. Including the characters of interest during tree reconstruction and the problems of circularity and bias in studies of character evolution. AmerNat. **148**:700-708.
- Desdevises, Y., P. Legendre, L. Azouzi, and S. Morand. 2003. Quantifying phylogenetically structured environmental variation. Evolution **57**:2647-2652.
- Diaz-Uriarte, R. & Garland, T. (1996) Testing hypotheses of correlated evolution using phylogenetically independent contrasts: sensitivity to deviations from Brownian motion. *Systematic Biology*, **45**, 27-47.
- Diniz-Filho, J. A. F., C. E. Ramos de Sant'Ana, and L. M. Bini. 1998. An eigenvector method for estimating phylogenetic inertia. Evolution 52:1247-1262.
- Estes S and Arnold SJ. 2007. Resolving the paradox of stasis: Models with stabilizing selection explain evolutionary divergence at all timescales. Amer. Nat. 169: 227-244.
- Farris, J. 1989. The retention index and the rescaled consistency index. Cladistics 5:417-419.
- Freckleton, R. P., P. H. Harvey, and M. Pagel. 2002. Phylogenetic analysis and comparative data: a test and review of the evidence. Amer. Nat. 160:712-726.
- Garland, T., Jr., P. H. Harvey, and A. R. Ives. 1992. Procedures for the analysis of comparative data using phylogenetically independent contrasts. Syst. Biol. **41**:18-32.
- Gittleman, J., and H.-K. Luh. 1992. On comparing comparative methods. Ann. Rev. Ecol. gittSyst. 23:383-404.
- Gittleman, J., and H.-K. Luh. 1994. Phylogeny, evolutionary models and comparative methods: a simulation study. Pages 103-122 *in* P. Eggleton and R. Vane-Wright, editors. Phylogenetics and ecology. Academic Press, London.
- Gittleman, J., and M. Kot. 1990. Adaptation: statistics and a null model for estimating phylogenetic effects. Syst.

Zool. 39:227-241.

- Givnish, T. 1997. Adaptive radiation and molecular systematics: issues and approaches. Pages 1-54 *in* T. J. Givnish and K. J. Sytsma, editors. Molecular evolution and adaptive radiation. Cambridge UnivPress, New York.
- Hansen, T. 1997. Stabilizing selection and the comparative analysis of adaptation. Evolution 51:1341-1351.
- Hansen, T. F., and S. H. Orzack. 2005. Assessing current adaptation and phylogenetic inertia as explanations of trait evolution: The need for controlled comparisons. Evolution 59:2063-2072.
- Harmon, L. J., J. A. Schulte II, A. Larson, and J. B. Losos. 2003. Tempo and mode of evolutionary radiation in Iguanian lizards. Science 301:961-964.
- Harvey, P. H., and T. H. Clutton-Brock. 1985. Life history variation in primates. Evolution 39:559-581.
- Harvey, P., A. Read, and S. Nee. 1995. Further remarks on the role of phylogeny in comparative ecology. JEcol. **83**:733-734.
- Harvey, P., A. Read, and S. Nee. 1995. Why ecologists need to be phylogenetically challenged. JEcol. 83:000-000.
- Kochmer, J., and S. Handel. 1986. Constraints and competition in the evolution of flowering phenology. EcolMonogr. **56**:303-325.
- Leroi, A. M., M. R. Rose, and G. V. Lauder. 1994. What does the comparative method reveal about adaptation? Amer. Nat. 143:381-402.
- Lord, J., M. Westoby, and M. Leishman. 1995. Seed size and phylogeny in six temperate floras: constraints, niche conservatism, and adaptation. Amer. Nat. **146**:349-364.
- Martins, E. 1996. Phylogenies, spatial autoregression, and the comparative method: a computer simulation test. Evolution **50**:1750-1765.
- McKitrick, M. C. 1993. Phylogenetic constraint in evolutionary theory: Has it any explanatory power? Ann. Rev. Ecol. Syst. 24:307-330.
- Morales, E. 2000. Estimating phylogenetic inertia in <u>Tithonia</u> (Asteraceae): A comparative approach. Evolution **54**:475-484.
- Orzack, S. H., and E. Sober. 2001. Adaptation, phylogenetic inertia, and the method of controlled comparisons. Pages 45-63 *in* S. H. Orzack and E. Sober, editors. Adaptationism and optimality. Cambridge University Press, Cambridge.
- Peterson, A. T., J. Soberon, and V. Sanchez-Cordero. 1999. Conservatism of ecological niches in evolutionary time. Science **285**:1265-1267.
- Rees, M. 1995. EC-PC comparative analyses. JEcol. 83:891-892.
- Revell, L. J., L. J. Harmon, and D. C. Collar. 2008. Phylogenetic signal, evolutionary process, and rate. Systematic Biology 57:591-601.
- Revell, L. J., L. J. Harmon, R. B. Langerhans, and J. J. Kolbe. 2007. A phylogenetic approach to determining the importance of constraint on phenotypic evolution in the neotropical lizard Anolis cristatellus. EVOLUTIONARY ECOLOGY RESEARCH 9:261-282.
- Revell, L. J., and L. J. Harmon. 2008. Testing quantitative genetic hypotheses about the evolutionary rate matrix for continuous characters. EVOLUTIONARY ECOLOGY RESEARCH 10:311-331.
- Ridley, M. 1992. Darwin sound on comparative method. TREE 7:37.
- Sanderson, M. 1991. In search of homoplastic tendencies: Statistical inference of topological patterns in homoplasy. Evolution **45**:351-358.
- Starck, J. 1998. Non-independence of data in biological comparisons a critical appraisal of current concepts assumptions and solutions. Theory Biosci. **117**:109-138.
- Wanntorp, H.-E. 1983. Historical constraints in adaptation theory: traits and non-traits. Oikos 41:157-160.
- Westoby, M., M. Leishman, and J. Lord. 1995. Further remarks on phylogenetic correction. JEcol. 83:727-729.
- Westoby, M., M. Leishman, and J. Lord. 1995. Issues of interpretation after relating comparative datasets to phylogeny. J. Ecol. **83**:892-893.
- Westoby, M., M. Leishman, and J. Lord. 1995. On misinterpreting the "phylogenetic correction". JEcol. 83:531-534.